Relationships between body condition and follicle development in mares¹

M.O. Gastal^{2,5}, E.L. Gastal³, V. Spinelli², O.J. Ginther⁴

Departments of Animal Science² and Veterinary³, Federal University of Viçosa, Viçosa, MG 36570, Brazil; Department of Animal Health and Biomedical Sciences⁴, University of Wisconsin, Madison, WI 53706, USA and Eutheria Foundation⁴, Cross Plains, WI 53528, USA

Abstract

Follicle activity and gonadotropin concentrations were compared between mares with low body condition (n=9) and mares with high body condition (n=8). Examinations began during the anovulatory season (August 14, Southern Hemisphere) and continued until the second ovulation of the year (63 to 141 days). Mares were fed with a complete diet of 1.5-2.0% of body weight in dry matter/day. Body condition increased slightly and similarly for the two groups during the study. Low body condition compared to high body condition was associated significantly with the following: longer interval to first ovulation (77.8 \pm 6.9 vs 63.0 \pm 3.8 days), smaller maximum diameter of the ovulatory follicle for the first ovulation $(45.6 \pm 1.4 \text{ vs } 51.1 \pm 1.0 \text{ mm})$ and second ovulation $(45.1 \pm 1.8 \text{ vs } 51.4 \pm 1.0 \text{ mm})$, fewer medium follicles (11-19 mm) per day preceding the first ovulation (6.0 \pm 0.0 vs 9.1 \pm 1.5) and fewer large follicles (\geq 20 mm) preceding the second ovulation $(1.3 \pm 0.2 \text{ vs } 2.0 \pm 0.2)$. During the last 19 days of the interovulatory interval, each of the four largest follicles was smaller in mares with low body condition than in mares with high body condition. There were no differences between groups in growth rate of the ovulatory follicle or in concentrations of FSH and LH, preceding either the first or second ovulations. Results indicated that low-body condition was associated with reduced follicle development, including diameter of the ovulatory follicle, during the transition between the anovulatory and ovulatory seasons and during the first interovulatory interval of the ovulatory season. These results were not attributable to

altered circulating concentrations of FSH and LH.

Keywords: body condition, follicles, mares.

Introduction

Inadequate nutrition or body condition has been associated with delayed onset of the breeding season, decreased pregnancy rate, increased embryo loss, and increased gestation length in mares (Henneke et al., 1983, 1984; Hines et al., 1987). During the winter, mares with low body condition had fewer medium (11 to 19 mm) and large (≥ 20 mm) follicles than mares with high body condition (Gentry et al., 2002). Apparently, the effects of inadequate nutrition or poor body condition on follicle dynamics during the equine ovulatory season are not known. In cattle, chronic or acute dietary restriction resulted in a gradual (Murphy et al., 1991; Bossis et al., 1999) or rapid (Mackey et al., 1999; Comin et al., 2002) reduction in growth rate and maximum diameter of the dominant follicle. Low dietary intake tended to increase the occurrence of estrous cycles with three major follicular waves in beef heifers (Murphy et al., 1991).

The mechanisms by which feed restriction modifies the reproductive axis are not well understood. Nutrients required for reproduction have not been differentiated from those required for other physiological functions. Initially, glucose, insulin, and fatty acids were considered as potential signals in the regulation of the reproductive axis; however, other studies have concluded that these metabolites do not play a role (Dowining *et al.*, 1995; Boukhliq *et al.*, 1996; Schreihofer *et al.*, 1996; Wade *et al.*, 1996).

¹ Supported by the Federal University of Viçosa, Minas Gerais State Foundation (FAPEMIG), Brazil and Eutheria Foundation, Cross Plains, Wisconsin, USA. Project P1-MOG-99. Part of these data were presented as a poster at the 33rd Annual SSR Meeting, Madison-WI.

⁵ Corresponding author: mgastal@eutheria.org, Fax 608-798-3722.

Received: June 30, 2004

Accepted: July 14, 2004

The present experiment was designed to study the relationship between body condition and follicle dynamics during the transition between the anovulatory and ovulatory seasons and during the first interovulatory interval in mares.

Materials and Methods

Animals and Groups

Twenty maiden, small draft-type, crossbred Breton mares, 3 to 4 years of age, and weighing 200 to 400 kg were used in the Southern Hemisphere (latitude, 21°). All mares were in the seasonal anovulatory phase characterized by follicles ≤ 20 mm and absence of a corpus luteum as determined by ultrasound examinations (Ginther, 1995). Mares were kept in corrals and fed daily with a diet consisting of forage (green grass, Pennisetum purpureum schum) and a mixture of grain, molasses and vitamins (Nutroeste; Nutrição Animal, Goiânia, GO, Brazil) to supply their maintenance requirements. Intake of dry matter per day ranged from 1.5 to 2.0% of body weight. Mares had free access to water and mineralized salt. The experiment was started during the last month of winter (August 14; equivalent to February in the Northern Hemisphere) and was concluded at the second ovulation of the ovulatory season. Mares that did not ovulate by 116 days after the beginning of the experiment were not used because of a requirement for normalizing to ovulation. Bodycondition score (1 to 9, lowest to highest; Henneke et al., 1983) and body weight were evaluated every 15 days. The score for body condition at each examination was determined by averaging the scores of two operators. At the first ovulation of the ovulatory season, the average of 4 to 6 body-condition scores for each mare was used to assign the mares to two groups: high body condition (score \geq 5; n=9) and low body condition (score <5; n=11).

Follicle Evaluations

An ultrasound scanner (Aloka SSD-500V; Aloka, Wallingford, CT) equipped with a 5 MHz linear-array transducer was used for transrectal ovarian examinations. Ovarian activity was evaluated over a span of 63 to 141 days, encompassing the first and second ovulations of the year; examinations were discontinued at the second ovulation in individual mares. Examinations were done every three days until a 25-mm follicle developed. After the largest follicle was \geq 25 mm, scanning was done daily until ovulation, using day-to-day identity of the largest follicle (Ginther, 1995).

Number of follicles per day was grouped into three categories: small (5-10 mm), medium (11-19 mm), and large (\geq 20 mm). Number of follicles was

assessed for 55 days before the first ovulation and 19 days before the second ovulation, using the mean value per mare over the indicated days in the comparison between groups. The number of major anovulatory waves per mare was taken from the beginning of the experiment to the first ovulation and between the first and second ovulations. A major anovulatory wave was identified by the presence of a nonovulatory follicle that reached \geq 30 mm (Ginther, 1993). Other follicle and gonadotropin end points were assessed for 31 days before the first ovulation and 19 days before the second ovulation. End points for the ovulatory follicles were diameter at maximum and at Day -1 (Day 0 =ovulation) and growth rate during the 7 days before ovulation. The four largest follicles were ranked as F1, F2, F3, and F4, according to descending diameter on each day of examination. The lengths of intervals from the beginning of the experiment to $a \ge 25$ -mm ovulatory follicle, from the beginning to the first ovulation, and from the first to the second ovulations (interovulatory interval) were also recorded.

Blood Samples and Hormone Assays

Jugular blood samples were collected every 3 days from each mare into heparinized tubes and centrifuged (500 xg for 10 min), decanted, and stored (-20 °C) until assay. Plasma samples were assayed for FSH and LH by radioimmunoassay as validated (Whitmore *et al.*, 1973; Freedman *et al.*, 1979, respectively) and modified (Donadeu & Ginther, 2002) in our laboratory. The intra- and interassay coefficients of variation and mean sensitivity were 12.7%, 11.7%, and 1.1 ng/mL for FSH and 8.4%, 5.0%, and 0.4 ng/mL for LH, respectively.

Statistical Analyses

Follicle data and plasma concentrations of gonadotropins were analyzed to determine effects of group, day and the interaction, using a mixed linear model with a repeated statement to account for the autocorrelation between sequential measurements (SAS, Institute Inc., Cary, NC). If a significant effect of group or group-by-day interaction was detected, unpaired t-tests were used to locate the mean differences between groups within a day, and paired t-tests were used between days within a group. The difference between groups in frequency of the number of mares with a major anovulatory wave was analyzed by chi-square. Pearson correlations between scores for body condition and maximum diameter of the ovulatory follicle were performed. A probability of $P \le 0.05$ indicated that a difference was significant. All data are expressed as the mean \pm SEM.

Results

Two mares from the low body-condition group did not ovulate by 116 days, resulting in nine mares in the group. A mare was removed from the high body-condition group because of an injury, resulting in eight mares in the group. Body-condition scores from the day at the beginning and the day at end of the experiment showed a main effect (P<0.0001) of group (high, 6.2 ± 0.3 ; low, 4.2 ± 0.1) and an effect (P<0.0006) of day (beginning, 4.6 ± 0.2 ; end, 5.5 ± 0.3), but the interaction was not significant. For body weight, there was an effect (P<0.0001) of group (high, 370 ± 12 ; low, 297 ± 12 kg) with no other significant differences.

Data associated with each of the two ovulations and the results of the statistical analyses are shown (Table 1). The mares with low versus high body condition had a longer interval from beginning of the

	Body condition ^a				
End points	High		Low		Probability
Intervals (days)					
Beginning of the experiment					
To ≥ 25 mm ovulatory follicle	50.4	± 3.6	68.8	± 6.8	P<0.02
To the first ovulation	63.0	± 3.8	77.8	± 6.9	P<0.04
25-mm ovulatory follicle to ovulation					
First ovulation	12.6	± 2.1	8.8	± 0.8	P<0.04
Second ovulation	10.3	± 0.8	9.9	± 0.9	NSb
First to second ovulation	22.8	± 1.1	24.0	± 2.6	NS
Ovulatory follicle					
Maximum diameter (mm)					
First ovulation	51.1	± 1.0	45.6	± 1.4	P<0.004
Second ovulation	51.4	± 1.0	45.1	± 1.8	P<0.003
Day -1^{c} diameter (mm)					
First ovulation	49.4	± 1.5	45.2	± 1.4	P<0.03
Second ovulation	51.1	± 1.0	44.3	± 1.4	P<0.001
Growth rate (mm/day) ^d					
First ovulation	1.3	± 0.4	1.8	± 0.3	NS
Second ovulation	2.5	± 0.4	1.9	± 0.6	NS
Number of follicles/day					
First ovulation					
5-10-mm	6.6	± 1.3	4.6	± 0.9	P<0.1
11-19-mm	9.1	± 1.5	6.0	± 0.0	P<0.05
\geq 20-mm	2.0	± 0.2	2.0	± 0.2	NS
Total (\geq 5 mm)	17.7	± 2.7	12.5	± 1.9	P<0.06
Second ovulation					
5-10-mm	5.9	± 0.7	5.2	± 0.9	NS
11-19-mm	5.8	± 0.8	4.0	± 1.0	P<0.09
\geq 20-mm	2.0	± 0.2	1.3	± 0.2	P<0.03
Total (\geq 5 mm)	14.0	± 1.5	10.0	± 1.7	P<0.05
Number of major anovulatory waves/mare ^e					
First ovulation	0.8	± 0.2	1.3	± 0.4	P<0.1
Second ovulation	0.4	± 0.2	0.4	± 0.2	NS

^aMares were grouped into those with high (≥ 5 ; n=8) versus low (<5; n=9) scores.

Anim. Reprod., v.1, n1., p.115-121, Oct./Dec. 2004

^bNS = not significant.

^cDay 0 =ovulation.

^dGrowth rate between Day -7 and -1.

^eLargest follicle \geq 30 mm.

experiment to a \geq 25-mm ovulatory follicle, longer interval to the first ovulation, and a shorter interval from a \geq 25-mm ovulatory follicle to ovulation. Diameter of the ovulatory follicle was smaller at the maximum diameter and at the day before the first and second ovulations in the low body-condition group. Diameter of the ovulatory follicle averaged over Days -7 to -1 before the first and second ovulations was larger (P<0.01) for the high-condition group (46.6 \pm 0.7 and 44.6 ± 0.8 mm, respectively) than for the lowcondition group (41.0 \pm 0.7 and 39.5 \pm 1.0 mm). There were fewer small and medium follicles per day before the first ovulation and fewer medium and large follicles before the second ovulation in the low- than in the high-condition group (Table 1). The total number of follicles (≥ 5 mm) per day was greater for mares with high body condition than for mares with low body condition before the first and second ovulations. A greater mean number of major anovulatory waves preceding the first ovulation in the low than in the high body-condition group approached significance. A greater number of mares with multiple major anovulatory waves in the low group (3/7) than in the high group (0/6) approached significance (P<0.07). For the mares that had one or more major anovulatory waves (7/9 and 6/8 in the low and high groups, respectively), there were more (P<0.04) waves per mare in the low group (1.4 ± 0.2) than in the high group (1.0 ± 0.0). The body-condition score was positively correlated with the ovulatory follicle at maximum diameter (P<0.01) and at Day -1 (P<0.0001) before the first ovulation (r=0.65 and r=0.59, respectively) and the second ovulation (r=0.83 and r=0.84).

During 31 days preceding the first ovulation, main effects of day indicated increasing diameters for F1 and F2 (Fig. 1). The group-by-day interaction was significant for only F3. There was no difference between groups or an interaction of group and day for either FSH or LH concentrations. A day effect for LH reflected increasing (P<0.05) concentrations between Days -7 to -1. During the 19 days preceding the second ovulation, there were main effects for both group and day but no interaction for any of the four follicles (Fig. 1). Although a day effect was obtained for both FSH and LH, there were no differences involving groups.



Figure 1. Mean \pm SEM diameter of the four largest follicles (F1, F2, F3, and F4) before the first ovulation of the year (left panel) and the second ovulation (right panel) in mares with high (solid lines) or low (broken lines) body-condition scores. The main effects and interactions that were significant or approached significance are shown. G = group effect; D = day effect; GD = group-by-day interaction. An asterisk indicates a difference (P<0.05) within a day, when a significant interaction was obtained.

Anim. Reprod., v.1, n1., p.115-121, Oct./Dec. 2004

Discussion

Body condition increased in the highcondition and low-condition groups during the experiment. There was no indication that the changes in body condition were different between groups (no day-bygroup interaction). The feeding program, slightly improved the initial body-condition scores and maintained body weights (no effect involving day) until the end of the experiment.

The longer interval from the beginning of the experiment to the first ovulation of the season in mares with low body condition is consistent with results of previous studies of feed restriction and/or poor body condition (Ginther 1979; Henneke et al., 1984; Kubiak et al., 1987). In addition, two mares in the lowcondition group were removed because they did not ovulate within 116 days; follicles attained diameters of 28 and 40 mm in these two mares. Although mares with low body condition needed more days for the ovulatory follicle to reach \geq 25-mm, the interval between a 25mm ovulatory follicle and ovulation was shorter but the growth rate of the ovulatory follicle after Day -7 was similar. These findings are consistent with the smaller diameter of the preovulatory follicle in the low bodycondition mares. Body condition did not alter the length of the first interovulatory interval of the reproductive season.

A negative effect of low body condition on the ovulatory follicle during the transition between the anovulatory and ovulatory seasons and during the first interovulatory interval was indicated by reduced diameters at maximum, on Day -1, and averaged over Days -7 to -1. These findings are consistent with the results of the correlation analysis which indicated that the greater the body condition, the greater the diameter of the ovulatory follicle at the maximum and at Day -1 for the first and second ovulations. Previous studies of the relationships between nutritional deficiency and follicle dynamics in cattle indicated negative effects on the maximum diameter and growth rate of the dominant follicle (Perry et al., 1991; Rhodes et al., 1996; Mackey et al., 1999). However, this is the first report of a negative effect of poor body condition on diameter of the first and second ovulatory follicles of the season in mares. In contrast to the cattle results, growth rate of the ovulatory follicle in mares was not altered by body condition.

Mares with low body condition had more multiple major anovulatory waves before the first ovulation of the year, although, in this regard, the interval from beginning of experiment to the first ovulation was 15 days longer in the low-condition group. Body condition may account for some of the differences among reports on the incidence of major anovulatory waves at the end of the anovulatory season in mares (Ginther, 1992), suggesting a need for further study. The occurrence of estrous cycles with three major follicular waves increased in beef heifers subjected to a low dietary intake (Murphy *et al.*, 1991), which seems similar to the present findings in mares.

Mares with low body condition had fewer medium follicles before the first ovulation of the season. This result has been reported previously in mares subjected to an inadequate nutrition (Van Niekerk and Van Heeden, 1972) or with low body condition (Gentry *et al.*, 2002) during the anovulatory season. However, fewer follicles during the ovulatory season (first interovulatory interval) in mares with poor body condition have not been previously reported.

Apparently this is the first report on the changing diameter of the four largest follicles during the transition between the anovulatory and ovulatory seasons and during the first interovulatory interval in mares with different body-condition scores. From Day -31 preceding the first ovulation, the main effect of day indicated increasing diameters of F1 and F2 for both body-condition groups. However, diameter of F1 was smaller in the low-condition group between Days -7 and -1. For the 19 days before the second ovulation, all four follicles were larger in mares with high body condition as indicated by the main effects of group. The day effect for each follicle indicated increasing diameter of F1 throughout the 19 days, contrasting with a plateau or decreasing diameters of F2-F4 after 10 days before ovulation. This result is consistent with the continuing increase in diameter of a dominant follicle (F1) and the decreasing diameters of subordinates follicles (F2-F4) after follicle deviation or selection during the estrous cycle (Ginther et al., 2003).

In the present study, the gonadotropins were not affected by body-condition. This finding agrees with previous studies in mares subjected to feedrestriction or with low body condition. Acute (McManus and Fitzgerald, 2000) or chronic (Gentry et al., 2002) feed restriction did not alter the plasma concentrations of LH, FSH, TSH, GH, glucose, or insulin. In contrast, in heifers (Rhodes et al., 1996), sheep (Miller et al., 1998), and pigs (Tokach et al., 1992) poor nutrition or body condition was associated with altered gonadotropins concentrations. Recently, the adipocyte-derived hormone leptin was suggested as a candidate for signaling nutrient status to the reproductive axis (Bray, 1996). A decrease in endogenous leptin secretions was temporally associated with cessation of reproductive activity during the anovulatory season in mares (Fitzgerald and McManus, 2000). It was proposed that, the reproductive response to a decrease in photoperiod or a presumptive inhibitory melatonin signal is modified by energy availability,

which may be signaled to the hypothalamus-pituitary axis through a change in the circulating concentration of leptin. In addition, low leptin, IGF-I, and prolactin plasma concentrations were observed in mares with low body-condition score during the middle of the anovulatory season (Gentry et al., 2002). Although the present study indicated a distinct effect of low body condition on the follicles associated with the first and second ovulations of the season, additional study will be required to elucidate the underlying mechanisms. Apparently, FSH and LH were not involved directly. Studies are needed in mares on the concentrations of follicular-fluid factors during low body condition. In this regard, the concentration of estradiol-17 β was reduced and the concentrations of NEFA (Nonsterified Fatty Acid), IGFBP2 and IGFBP3 (IGF - Binding Protein) were increased in the follicular fluid of dairy cows subjected to feed restriction (Comin et al., 2002).

In conclusion, results of this study confirmed previous findings that poor body condition during the transition between the equine anovulatory and ovulatory seasons is associated with a delayed beginning of the ovulatory season. Novel findings were that body condition affected development of the ovulatory follicle for both the first and second ovulations of the ovulatory season, as shown by smaller diameter of the follicle in mares with low body condition. In addition, mares with low body condition had fewer follicles ≥ 5 mm before both the first and second ovulations. In the low body-condition group, only the largest follicle was negatively affected (Days -7 to -1) before the first ovulation; however, the diameter of the four largest follicles was reduced during 19 days before the second ovulation.

Acknowledgments

The authors thank Nutroeste (Nutrição Animal, Goiânia, GO, Brazil) for a gift of the grain mixture, and Ana Paula Gonçalves Mellagi, Thiago Hollanda Ayup, and Fernando Antônio de Freitas for care and handling of the animals.

References

Bray GA. 1996. Leptin and leptinomania. *Lancet*, 348-140-141.

Bossis I, Welty SD, Wettemann RP, Vizcarra JA, Spicer LJ, Diskin MG. 1999. Nutritionally induced anovulation in beef heifers: ovarian and endocrine function preceding cessation of ovulation. *J Anim Sci*, 77:1536-1546.

Boukhliq R, Miller DW, Martin GB. 1996. Relationship between nutritional stimulation of gonadotropin secretion and the peripheral and cerebrospinal fluid (CSF) concentrations of glucose and insulin in rams. *Anim Reprod Sci*, 41:201-214.

Comin A, Gerin D, Cappa A, Marchi V, Renaville R, Motta M, Fazzini U, Prandi A. 2002. The effect of an acute energy deficit on the hormone profile of dominant follicles in dairy cows. *Theriogenology*, 58:899-910.

Donadeu FX, Ginther OJ. 2002. Changes in concentrations of follicular fluid factors during follicle selection in mares. *Biol Reprod*, 66:1111-1118.

Dowining JA, Joss J, Scaramuzzi RJ. 1995. Ovulation rate and the concentrations of gonadotrophins and metabolic hormones in ewes infused with glucose during the late phase of the oestrous cycle. *J Endocrin*, 146:403-410.

Fitzgerald BP, **McManus CJ.** 2000. Photoperiodic versus metabolic signals as determinants of seasonal anestrus in the mare. *Biol Reprod*, 63:335-340.

Freedman LJ, Garcia MC, Ginther OJ. 1979. Influence of photoperiod and ovaries on seasonal reproductive activity in mares. *Biol Reprod*, 20:567-574.

Ginther OJ. 1979. 1.ed. *Reproductive biology of the mare: basic and applied aspects*. Cross Plains, WI, USA: Equiservices Publishing.

Ginther OJ. 1992. *Reproductive biology of the mare: basic and applied aspects.* 2nd.ed. Cross Plains, WI, USA: Equiservices Publishing.

Ginther OJ. 1993. Major and minor follicular waves during the equine estrous cycle. *J Equine Vet Sci*, 13:18-25.

Ginther OJ. 1995. Ultrasonic imaging and animal reproduction: Book 2, Horses. Cross Plains, WI, USA: Equiservices Publishing.

Ginther OJ, Beg MA, Donadeu FX, Bergfelt DR. 2003. Mechanism of follicle deviation in monovular farm species. *Anim Reprod Sci*, 78:239-257.

Gentry LR, Thompson Jr. DL, Gentry Jr. GT, Davis KA, Godke RA, Cartmill JA. 2002. The relationship between body condition, leptin, and reproductive and hormonal characteristics of mares during the seasonal anovulatory period. *J Anim Sci*, 80:2695-2703.

Henneke DR, Potter GD, Kreider JL. 1984. Body condition during pregnancy and lactation and reproductive efficiency of mares. *Theriogenology*, 21:897-909.

Henneke DR, Potter GD, Kreider JL, Yeats BF. 1983. Relationship between body condition score, physical measurements and body fat percentage in mares. *Equine Vet J*, 15:371-372.

Hines KK, Hodge SL, Kreider JL, Potter GD, Harms PG. 1987. Relationship between body condition and levels of serum luteinizing hormone in postpartum mares. *Theriogenology*, 28:815-825.

Kubiak JR, Crawford BH, Squires EL, Wrigley RH, Ward GM. 1987. The influence of energy intake

and percentage of body fat on the reproductive performance of nonpregnant mares. *Theriogenology*, 28:587-598.

Mackey DR, Joseph MS, Roche JF, Diskin MG. 1999. Effect of acute nutritional restriction on incidence of anovulation and periovulatory estradiol and gonadotropin concentrations in beef heifers. *Biol Reprod*, 61:1601-1607.

McManus CJ, Fitzgerald BP. 2000. Effects of a single day of feed restriction on changes in serum leptin, gonadotropins, prolactin and metabolites in aged and young mares. *Domest Anim Endocr*, 19:1-13.

Miller DW, Blache D, Boukhliq R, Curlewis, JD, Martin GB. 1998. Central metabolic messengers and effects of nutrition on gonadotrophin secretion in sheep. *J Reprod Fertil*, 112:347-356.

Murphy MG, Enright WJ, Crowe MA, McConnell K, Spicer LJ, Boland MP, Roche JF. 1991. Effect of dietary intake on pattern of growth of dominant follicles during the oestrus cycle in beef heifers. *J Reprod Fertil*, 92:333-338.

Perry RC, Corah LR, Cochran RC, Beal WE, Stevenson JS, Minton JE, Simms DD, Brethour JR. 1991. Influence of dietary energy on follicular development, serum gonadotropins, and first postpartum

ovulation in suckled beef cows. *J Anim Sci*, 69: 3762-3773.

Rhodes FM, Entwistle KW, Kinder JE. 1996. Changes in ovarian function and gonadotrophin secretion preceding the onset of nutritionally induced anoestrus in *Bos indicus* heifers. *Biol Reprod*, 55:1437-1443.

Schreihofer DA, Renda F, Cameron JL. 1996. Feeding-induced stimulation of luteinizing hormone secretion in male rhesus monkeys in not dependent on a rise in blood glucose concentration. *Endocrinology*, 137:3770-3776.

Tokach MD, Pettigrew JE, Dial GD, Wheaton JE, Crooker BA, Johnson LJ. 1992. Characterization of luteinizing hormone secretion in the primiparous, lactating sow: relationship to blood metabolites and return-to-estrus interval. *J Anim Sci*, 70:2195-2201.

Van Niekerk CH, Van Heeden JS. 1972. Nutrition and ovarian activity of mares early in the breeding season. *J S Afr Vet Med Assoc*, 45:351-360.

Wade GN, Schneider JE, Li H-Y. 1996. Control of fertility by metabolic cues. *Am J Physiol*, 270:E1-E19.

Whitmore HL, Wentworth BC, Ginther OJ. 1973. Circulating concentrations of luteinizing hormone during estrous cycle of mares as determined by radioimmunoassay. *Am J Vet Res*, 34:631-636.